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REVIEW

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Anthocyanins: Plant-based flavonoid pigments with diverse biological activities

Sandip Patra^a, Priyanka N. Makhal^a, Shubham Jaryal^b, Nilesh More^a, Venkata Rao Kaki^{a,*}

^a National Institute of Pharmaceutical Education and Research (NIPER), Department of Medicinal Chemistry, Hyderabad 500037, India ^b Ind-Swift Laboratories Limited, Mohali, Punjab 160055, India

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1. Introduction

The term Anthocyan has been derived from the Greek word Anthos, which means flower, and kyanos means blue (Marquart, 1835). It was introduced by Ludwig Clamor Marquart, a German pharmacist, to distinguish the blue color pigments of various flowers. In the plant kingdom, most species contain anthocyanins (Richardson and Harborne, 1990). They exist in all tissues, including leaves, stems, fruits, flowers, and roots of the higher plants. Anthocyanins are found in the cell vacuole, which gives color shade to the plant tissue such as red, blue, and purple as per their essential structure organization. Fruit plants such as red raspberry, blackberry, bilberry, blueberry, grapes, zante currants, cherry, blood orange, red cabbage, red onion, purple sweet potato, red-skinned potato, fennel, eggplant, and radish can be found in these plants (Table 1).

* Corresponding author: E-mail address: kvenkata.rao@niperhyd.ac.in (V.R. Kaki) e-ISSN: 2791-7509

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ABSTRACT

Anthocyanins are flavonoid containing polyphenolic phytochemicals. They are widely present in plants and accounts for different color shades displayed by the plant organs. A broad range of health-revitalizing effects is attributed to anthocyanins, constituting a vital part of the human diet. They are also accountable for ameliorating the detrimental effects of various lifestyle diseases, including cancer, cardiovascular disorders, neurological disorders, etc. These beneficial impacts highly depend on the bioavailability of anthocyanins, governed by their absorption and metabolism in the human body. The primary goal of this review is to summarize the latest anthocyanin knowledge while focusing on the chemistry, pharmacokinetics, and various biological advantages with anti-cancer, neuroprotective, antidiabetic, antioxidant, cardiovascular protective, vision improvement, antiviral, and antimicrobial effects.

Anthocyanins play a shielding role in vegetables and plants, including monosaccharide transport vehicles, the absorbance of harmful radiations, and osmotic adjuster during frost or drought (Clifford, 2000). These also play a critical role in attracting livestock facilitating pollination and dispersal of seeds (Eder, 2000; Timberlake and Henry, 1988).

Recently, anthocyanins have been the primary focus of researchers for their pharmacological effects, biological properties, and highwater solubility. Certain nutritionists are progressively suggesting the utilization of foods containing anthocyanins. They are less active compared with related drugs but show physiological results of great importance when consumed in a regular diet. In traditional medicinal items, several red flowers (clove, hibiscus, pineapple sage, and rose), purple (passionflower, lavender, and purple sage), and blue (blue rosemary and cornflower) are used (Calderaro et al., 2020).

A few exceptional reports have delineated these natural dietary phytochemicals in terms of their intake (McGhie and Walton, 2007), metabolism (He and Giusti, 2010), pharmacokinetics (Kay, 2006),

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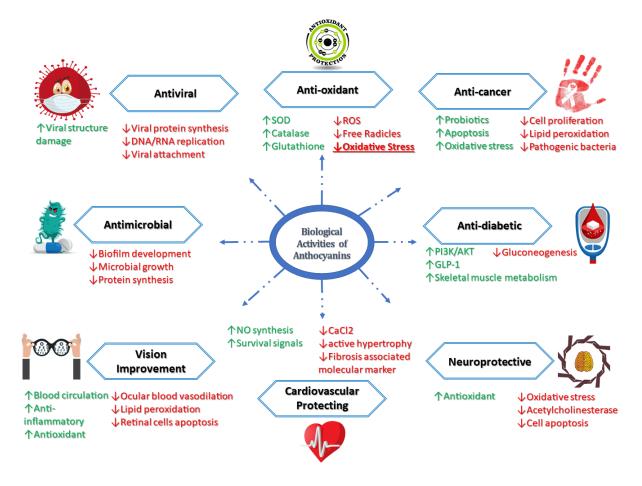
molecular mechanisms (Li et al., 2017), and bioavailability (Fernandes et al., 2014) over the past few years. In recent years, the enthusiasm for studying anthocyanins' health and medical benefits has increased rapidly. Vanamala (2019) has summarized the effect of red and purple anthocyanins on colon cancer stem cells. Khan et

al. (2019) have checked out the neuroprotective effect of anthocyanins, whereas Krga and Milenkovic (2019) have described the impact of anthocyanins on cardiovascular diseases.

Table 1. Total anthocyanin content in common fruits and vegetables (Clifford, 2000; Eder, 2000; Timberlake and Henry, 1988)

Fruit or vegetable	Total anthocyanin content (mg/kg)		
Cherry	3500-4500		
Cranberry	460-2000		
Strawberry	127-360		
Grapes (blue)	80-3880		
Orange	2000		
Grapes (red)	300-7500		
Plum	19-250		
Cabbage (red)	250		
Onion (red)	Up to 250		
Bilberry	4600		
Eggplant	7500		
Elderberry	2000-15600		
Radish (red)	110-600		

This review describes the chemistry of anthocyanins and recent literature about various biological benefits of anthocyanins, such as anti-cancer activity, neuroprotective activity, anti-diabetic, antioxidant, cardiovascular protecting activity, vision improvement, antiviral and antimicrobial effects (Figure 1).





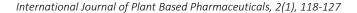
2. Chemistry of anthocyanins and anthocyanidins

Anthocyanins are anthocyanidin glycosides, have a distinctive carbon skeleton with 2 phenyl rings (A and B rings, Figure 2) and a heterocyclic pyran (C ring) fused with A ring (Castañeda-Ovando et al., 2009). Cytoplasmically-localized glycosyltransferases generally catalyze glycosylation, and the most common sugars are L-

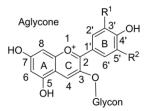
arabinose, L-rhamnose, D-galactose, D-glucose, and D-xylose. Sugars are usually bound to the 3-hydroxyl position of anthocyanidins or the 5- or 7- position of anthocyanidins (Pina et al., 2012). Anthocyanidins (aglycone) are chemically flavylium salts or (2-phenylchromenylium) ion compounds. With a molecular weight of 207.25, the molecular formula for the flavylium ion is $C_{15}H_{11}O^+$. The biosynthetic process of anthocyanins is initiated from the acetate

and shikimate pathways. Anthocyanins are transported into the vacuole by 3 possible mechanisms after completion of their biosynthesis: (i) via vesicle-mediated mass transport; (ii) via glutathione S-transferase-like protein; and (iii) via flavonoid/H⁺ antiporter (Zhang et al., 2006). In the vacuole, the anthocyanins occur as the equilibrium of four molecular species, are essentially flavylium cation (I), the quinoidal base (II), the carbinol pseudo base (III), and the chalcone pseudo base (IV) (called as secondary structures, Figure 3) (Prior and Wu, 2006).

Over 600 anthocyanins have been identified, and the number continues to increase exponentially (Kong et al., 2003). Anthocyanins can be distinguished based on (i) the number and location of the hydroxyl group, (ii) the degree of hydroxyl group



methylation, (iii) the number and nature of sugar (glycon) molecules linked to the aglycone portion, and (iv) the acylation of the sugar (aliphatic or aromatic acid) molecules. Caffeoylated anthocyanin, malonylated anthocyanin, acrylated anthocyanin, and coumaroylated anthocyanin can be further categorized as the acylated form of anthocyanins. Both color and hue of anthocyanins are affected by hydroxylation, glycosylation, methylation, and acylation pattern of the phenyl constituent of the anthocyanidins, and the color varies from orange-red (cyanidin, pelargonidin) to purple and pink-magenta (petunidin, delphinidin, malvidin, and peonidin) (da Costa et al., 2000; Iversen, 1996; Keppler and Humpf, 2005; Khoo et al., 2017; Lo Piero, 2015).



Glycon = D-glucose, D-galactose, D-xylose L-rhamnose, L-arabinose

S. No.	Anthocyanidin (or aglycone)	R1	R ²	Distribution in fruits and vegetables (%)	Major metabolic degraded products
1	Cyanidin	OH	Н	50	Protocatechuic acid
2	Delphinidin	OH	OH	12	Gallic acid
3	Pelargonidin	Н	Н	12	4-Hydroxybenzoic acid
4	Petunidin	OCH₃	OH	7	3-O-methylgallic acid
5	Lanvidin	OCH ₃	OCH₃	7	Syringic acid
6	Peonidin	OCH ₃	н	12	Vanillic acid

Figure 2. Anthocyanidins relevance to human diet - general structure, distribution, and metabolic products (Leong et al., 2018; Turturică et al., 2015)

It is currently thought that six anthocyanins are relevant to the human diet (Figure 2). Their distribution in fruits and vegetables are 50% cyanidin, 12% delphinidin, 12% pelargonidin, 12% peonidin, 7% malvidin, and 7% petunidin (Leong et al., 2018; Turturică et al., 2015).

Due to the presence of electron-deficient flavylium cation, anthocyanins are sensitive and highly reactive, and thus easier to be degraded (Felgines et al., 2007; Yang et al., 2011). Anthocyanin's stability is affected by its molecular structure. Due to the reactive - OH group; delphinidin and cyanidin are considered more stable than peonidin, malvidin, and petunidin. Different transformations at position 4 resulting in hetero-dimers, condensation, and cycloaddition products, make anthocyanins more durable. Glycosylation at position 3 makes it stable, whereas glycosylation at position 5 leads to instability. The stability of anthocyanins increases with acylation by intramolecular pigmentation and self-association reactions (Clifford, 2000).

Since the anthocyanins' molecular structure has an ionic nature, the solution's pH determines the anthocyanins' color (Fossen et al., 1998). In acidic solution (pH 1-3), some of the anthocyanins appear red due to flavium cation formation (I), whereas in neutral pH, they appear colorless due to carbinol pseudo base formation (III). At the same time, color changes from blue to purple as pH increases to 7-8 due to the quinoidal base formation (IV) (Figure 3) (Hribar and Poklar Ulrih, 2014). It is determined that the stability of anthocyanins is directly proportional to the density of their solutions, but intermolecular interaction within the formulation

often improves balance by hindering water-based anthocyanin deterioration.

3. Absorption of anthocyanins

In recent years, the study of dietary intake and health-promoting effects of anthocyanins has been increased. The aiding and therapeutic benefits of anthocyanins directly depend on the pharmacokinetic properties (absorption, distribution, metabolism, and excretion) and consequently bioavailability of individuals. The absorption mainly relies on the chemical structure, nutrient components of food, gut microbiota, and individual genetic factors (Felgines et al., 2007; Yang et al., 2011). For instance, anthocyanins based on pelargonidin or 3'-hydroxyanthocyanins are more quickly absorbed than other anthocyanins with more B-ring substituents. Usually, non-acylated anthocyanins show more absorbance capacity as compared to acylated anthocyanins.

The absorption of anthocyanin occurs within the stomach and mainly occurs in the small intestine, although it is absent in the intestinal tract. Bilitranslocase, an organic anion membrane transporter located in the gastric mucosa, produces gastric absorption (Passamonti et al., 2002). Injection of red grape wine anthocyanins into the stomach of Wistar male rats, followed by the blood from the portal vein analysis, confirmed the existence of malvidin-3-O-glucoside in the heart (Passamonti et al., 2003). Similarly, hexose transporters expressed on intestinal epithelium are accountable for anthocyanins' active and passive transport (Han et al., 2019). Talavéra et al. (2004) used an *in situ* perfusion procedure to examine the absorption of anthocyanins in the small intestinal

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tract. The rate of anthocyanins absorption in the small intestine was determined by measuring the number of anthocyanins left in the effluent; the absorption rate for cyanidin-3-*O*-glucoside was found to be 22%, and for malvidin-3-glucoside, it was found to be 10-11%. He et al. (2009) determined that 7.5% of the black raspberry anthocyanins administered were absorbed into the small intestinal tissues of rats. In line with the above predictions, animal model studies also demonstrate that anthocyanins quickly appear in the

systemic circulation (6-20 minutes) and reach a maximum blood level after 15-50 minutes (Pojer et al., 2013). However, few experimental studies demonstrate contradictory results, presenting that there is a minimal quantity of anthocyanins (i.e., < 1% of the consumed amount) in systemic blood circulation or the plasma (Prior, 2004).

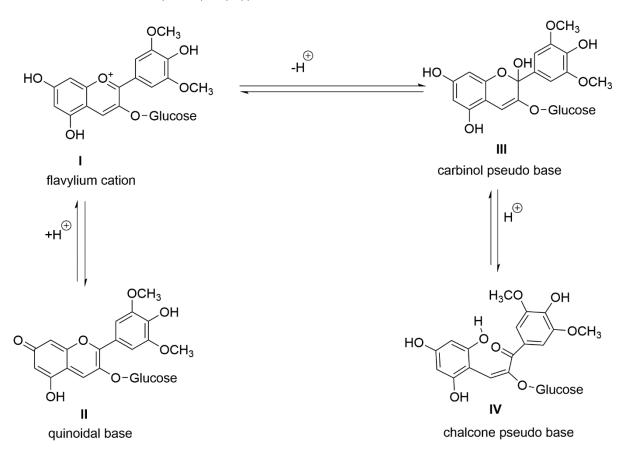


Figure 3. Anthocyanins equilibria (Prior and Wu, 2006)

Temperature, intestinal pH, and the microbes influence the digestion of anthocyanins in the human intestinal tract (Hidalgo et al., 2012). Breaking the glycosidic linkage among the sugar and aglycone moieties is activated by intestinal microflora and releases aglycone. The microbes responsible for the metabolism include Clostridium, Bacteroides, Eggertheilla, Ruminococcus, and Eubacterium (Blaut and Clavel, 2007; De Ferrars et al., 2014; Jang and Kim, 1996). The basic conditions and microbiota further convert aglycons into phenolic aldehydes and acids (Hanske et al., 2013). When cyanidin-3-glucoside was incubated with the intestinal bacteria, a critical aggregation of metabolic degradation products including 2,4,6-trihydroxy benzaldehyde and protocatechuic acid was brought about. The reported primary metabolites from anthocyanins relevant to the human diet are protocatechuic acid, 3-O-methyl gallic acid, 3,5-dihydroxybenzoic acid, vanillic acid, and syringic acid contingent upon the parent anthocyanin structure (Figure 2) (Muñoz-González et al., 2013; Nurmi et al., 2009). The usual chemical transformations such as demethylation, hydrogenation, hydroxylation, demethoxylation, decarboxylation, hydrolysis, and dehydroxylation occur during the degradation of anthocyanins and their metabolites (Forester and Waterhouse, 2008; Stalmach et al., 2013).

4. Biological activities of anthocyanins

4.1. Antioxidant effects

Compounds that are easier to oxidize are often termed antioxidants. They can also be described as molecules, giving free hydrogen or electron atom to react with free radicals. Several therapeutic activities of anthocyanins are mainly contributed by their antioxidant property (Joshi et al., 2017; Lucioli, 2012). Anthocyanins' free radical scavenging property confers a superior antioxidant activity. The antioxidant activity of anthocyanins relay upon its structure, and the properties are influenced by (i) the B ring catechol moiety; (ii) the number of -OH group; (iii) the hydroxylation and methylation pattern; (iv) acylation; (v) glycosylation; and (vi) the oxonium ion present in the C ring. Glycosylation of anthocyanins lessens the antioxidant properties due to the inability of anthocyanin radicals to delocalize its electron. Acylation of anthocyanins with phenolic acids shows a prominent result of increased antioxidant activities. Diacylation of anthocyanins also shows increased antioxidant activity, whereas 5-glycosylation minimizes its activity. When comparing anthocyanins with oligomeric proanthocyanidins and other flavonoids, anthocyanins' antioxidant strength and hydrogen-donating effect are greatly

affected by the positive charged O_2 atom in the scaffold (Shaik et al., 2018; Smeriglio et al., 2016). There may be two mechanisms by which anthocyanins impose their antioxidant effect, i.e., direct and indirect pathways. These exhibit direct free radical scavenging behavior due to the hydrogen (electron) donating potential of the flavonoid molecules, which can form a bond with reactive oxygen species (ROS) such as singlet oxygen $({}^{1}O_{2})$, superoxide (O_{2}) , hydroxyl radical species (OH), and hydrogen peroxide (H₂O₂) (Borkowski et al., 2005; Janeiro and Brett, 2007; Tsuda et al., 1996). On the other hand, anthocyanins increase the antioxidant property through various mechanisms such as: (i) increasing or restoring the superoxide dismutase (SOD), glutathione peroxidase, and the antioxidant enzymes, consequently elevating glutathione content; (ii) decreased production of DNA oxidative adducts, decreased production of endogenous ROS by obstructing xanthine oxidase and NADPH oxidase, or by altering the arachidonic metabolism and respiration of mitochondria; and (iii) activation of genes that code for the above antioxidant enzymes.

Anthocyanins-rich food mostly contains cyanidin, malvidin, peonidin, and these all play a crucial role in scavenging radicals and protecting them from DNA damage. For example, eggplant shows antioxidant activity by scavenging the oxygen-free radical with > 65% scavenging activity achieved in 2,2-diphenylpicrylhydrazyl (DPPH) assay (Li et al., 2012). Similarly, sweet cherry (Prunus avium L.) anthocyanins demonstrate antioxidant properties with above 70% activity on the DPPH (Sonmez et al., 2022). Likewise, blackberries exhibit the scavenging action against superoxide radicals, and the anthocyanins present in red cabbage protect it from oxidative stress (Igarashi et al., 2000; Murapa et al., 2012; Wang and Jiao, 2000; Zafra-Stone et al., 2007). Delphinidin-3-glucoside (IC $_{50}$ values: 1.6 and 0.7 $\mu M)$ and delphinidin (IC₅₀ values: 2.6 and 0.9 µM) shows maximum inhibitory effect against ¹O₂ scavenging activity and lipid peroxidation, respectively, whereas pelargonidin has the greatest inhibitory effect on the -OH radical scavenging activity with IC_{50} value of 8.5 μM (Tsuda et al., 1996). The presence of 3-OH groups on the B ring advances the antioxidant properties of delphinidin. However, oxidation of the hydroxyl group can be achieved by donating one electron (semiguinone form) or two-electron (guinone form).

4.2. Anti-cancer activity

Anthocyanins' chemopreventive effects are largely due to their antiinflammatory and antioxidant activities. The consumption of anthocyanin-rich foods and vegetables leads to anti-cancer effects through different mechanisms like (i) anti-angiogenesis and inducing apoptosis, (ii) prohibiting oxidative DNA damage, (iii) inhibiting COX-2 enzymes (iv) inhibition of cell proliferation. 2.5-100 µM extracts from anthocyanin-containing berries of different types and eggplant and black rice tend to inhibit the invasion of cancer cell types in Matrigel (Wang and Stoner, 2008). Strong anti-proliferative activity was shown by the hydrolyzed pulp of Jamun with an IC₅₀ value of 59 \pm 4 µg/ml. Anthocyanins extracted from berries, especially black raspberries, help lower the epigenetic factors involved in cancer. Histone modification and DNA methylation are part of this epigenetic regulation of gene expression. They are involved in the demethylation of tumor suppressor genes and the suppression of DNA methyl transferase and thus help prevent cancer (El-Ella and Bishayee, 2019). Studies of anthocyanin extracts in vivo and in vitro (cancer cells) have shown anti-carcinogenic effects on breast cancer (Singletary et al., 2007), prostate and liver cancer (El-Ella and Bishayee, 2019; Lin et al., 2017), colon cancer (Jaganathan et al., 2014) and lung cancer. Anthocyanin rich extract of Syzygium cumini (Aqil et al., 2012), purple-fleshed sweet potato (clone P40) (Lim et al., 2013), raspberries, strawberry (Giampieri et al., 2018; Zhang et

al., 2008), sorghum, black lentil, and red grapes exhibited antiproliferative action on the cancer cells (Grimes et al., 2018; Mazewski et al., 2018). Anthocyanidins seem to inhibit different kinase pathways and show better antiproliferative activity than anthocyanins (Zhang et al., 2005). Delphinidin-3-glucoside and cyanidine-3-glucoside inhibit the development of colon cancer HT-29 cells (Grimes et al., 2018). Delphinidin has the best inhibitory characteristics as it possesses a hydroxyl group in its structure, which seems to show its effect by halting the MAPK pathway (Hou, 2003). Anthocyanins, gallic acid, 3-O-methyl gallic acid, and 2,4,6trihydroxybenzaldehyde metabolites reduce cell viability and apoptosis of colon cells in Caco-2 cells (Forester et al., 2014). A recent study shows that the petal anthocyanins from Crocus sativus (saffron) have a beneficial effect on the mouse model with polycystic ovary syndrome. Saffron petal anthocyanins hinder the polycystic ovary syndrome by improving antioxidant enzymes, steroidogenic dysregulation of ovarian steroids, and inflammatory markers (Moshfegh et al., 2022). Likewise, it was found that the total anthocyanins extracted from sweet cherry (Prunus avium L.) inhibit human carbonic anhydrase I and II enzyme activities and thus show the anticancer effect (Sonmez et al., 2022).

4.3. Anti-diabetic activity

Diabetes (Type-2) mainly emerges because of continuous stress on the pancreas, β -cells degeneration, decreased insulin level, and expanded insulin resistance. In hyperglycemic and hyperlipidemic states, there is high oxidative stress upon pancreatic β -cells, which may lead to the execution of these cells. Consumption of anthocyanins can reduce free radicals' excessive production, which prevents the β -cells from oxidative damage (Al-Awwadi et al., 2005; Cao et al., 2019; Hossain et al., 2016). Anthocyanins inhibit alphaglucosidase, an enzyme responsible for the hydrolysis of small intestinal carbohydrates that helps control diabetes (Zhang et al., 2019). Similarly, the anti-inflammatory effects of anthocyanins manifested by decreasing monocyte chemotactic protein-1 (MCP-1), tumor necrosis factor-alpha (TNF-alpha), interleukin-6 (IL-6) expression help in the recovery of insulin resistance and type 2 diabetes (Khanra et al., 2015; Zhu et al., 2012). Infusion of pelargonidin on streptozotocin (STZ)-induced diabetes rats brought about an increased serum level of superoxide dismutase and lowered the level of malondialdehyde and fructosamine (Roy et al., 2008). Cyanidin, a major anthocyanidin present in plants, vegetables, and fruits, helps in inciting insulin secretion by raising the level of intracellular calcium in pancreatic β -cells (Pesce and Menini, 2019). Zhu et al. (2012) examined the effect of cyanidin-3glucoside on HepG2 cells and was found to increase the expression of glutamate-cysteine ligase, which reduces the level of ROS. Black soybean seed coat extract contains peonidin-3-O-glucoside, delphinidin-3-O-glucoside, and cyanidin-3-O-glucoside, showing protection against obesity and diabetes, reducing oxidative stress, inflammation, level of α -amylase, and lipid aggregation in adipocytes in HepG2 cells. Fasting blood glucose (FBG) and oral glucose tolerance test (OGTT) results obtained in diabetic mice showed black soybean seed coat extract (BSSCE) to exhibit the hypoglycemic effect. BSSCE could alleviate FBG levels and cause a significant decrease (p < 0.05) in the AUC value concerned with OGTT at a concentration of 400 and 200 mg/kg, respectively, which was comparable to positive control drug rosiglitazone (Chen et al., 2018).

4.4. Neuroprotective effects

Studies show that anthocyanins defend against several neurodegenerative diseases, including Parkinson's disease,

Alzheimer's disease, ischemia, and other neuronal injuries (Airoldi et al., 2018; Jung and Kim, 2018). The glycosylated forms of anthocyanins could cross the blood-brain barrier and reach the CNS to convey their natural and biological impacts. Collective evidence suggests that the transfer of anthocyanin takes place through a bilitranslocase transporter, mainly into the vascular endothelium and thus into the target tissue (Aqil et al., 2014; Maestro et al., 2010). The polyphenolic cationic structure of anthocyanins allows scavenging free radicals, reducing ROS formation, giving an impression of being profoundly viable in neurodegenerative diseases. Anthocyanins help boost the PI3K/Akt/GSK3 pathway and control the endogenous antioxidant Nrf2/HO-1 pathway in Alzheimer's disease. Anthocyanin (12mg/kg i.p. for 30 days) significantly improved memory function in APP/PS1 transgenic AD mice (Ali et al., 2018). The arrangement of soluble amyloid- β 25-35 oligomers and their neurotoxicity in the human neuronal cell line SH-SY5YY could be prevented by cyanidin-3-O-glucoside (Tarozzi et al., 2010). Furthermore, if malvidin-3-O-glucoside is tested with amyloid- β neuronal cells, it shows inhibition of the cell cycle interruption caused by the amyloid- β (Shih et al., 2011). An *in silico* study found that anthocyanins could cause conformational modifications that trigger FKPB52, a potential protein known for restricting tau accumulation (Cao and Konsolaki, 2011).

Likewise, anthocyanins inhibit inflammatory biomarker interleukin-8 and other pro-inflammatory cytokines, which may help curb neuronal apoptosis associated with the disease (Rasheed et al., 2009). When anthocyanins-rich extract of blueberry are treated with microglial cells, it reduces the expression of iNOX and COX genes, which are involved in the inflammatory process (Bensalem et al., 2015). Anthocyanins inhibit extracellular signal-regulated kinase (ERK), mitogen-activated protein kinases (MAPKs), and c-Jun Nterminal kinase (JNK), which are essential for the expression of proinflammatory cytokines (Bensalem et al., 2015; Pan et al., 2018; Rasheed et al., 2009; Shah et al., 2016).

4.5. Cardiovascular protecting effects

Cardiovascular disease is the leading cause of mortality globally and is the type of disease affecting the cardiovascular system. Several physiological regulations lead to cardiovascular disorder, including high plasma LDL cholesterol, hypertension, endothelium dysfunction, and platelet aggregation. Apart from all these, oxidative stress is one of the major risk factors for cardiovascular disorders. Anthocyanins play a vital role in protection against oxidative stress (Bell and Gochenaur, 2006). Delphinidin, which is present abundantly in red wine, shows a vasodilatory effect upon consumption. However, in terms of preventing atherosclerosis and improving lipid and antioxidant parameters, the juice of red grapes was more efficacious than red wine (Andriambeloson et al., 1998). Literature suggests that during a heart attack, ingestion of grape juice, which is rich in anthocyanins, could exhibit potent antioxidant activity by increasing capillary permeability and strength. This action also leads to inhibition of platelet formation and speed-up the nitric oxide (NO) production, which results in vasodilation (Andriambeloson et al., 1998; Erlund et al., 2008). Similarly, chokeberry extracts also possess vasorelaxation properties (Bell and Gochenaur, 2006; Bertuglia et al., 1995). Clinical trial studies have shown that in healthy participants, eating anthocyanin-rich strawberry for more than a month improves the lipid profile and platelet function (Alvarez-Suarez et al., 2014). Hyperlipidemia is known to be an important cardiovascular disease risk factor. The literature study shows that black currant, rich in anthocyanin, can act against the fatty acids to reduce its levels (Frank et al., 2002). Anthocyanins can carry out their cardiovascular protective action

through their anti-inflammatory properties; they inhibit inflammation caused by TNF- α in the human endothelium via monocyte chemoattractant protein-1 (Garcia-Alonso et al., 2009). Free radicals are formed during ischemia-reperfusion injury, which leads to adhesion of WBC in the wall of the micro-capillary; thus, it reduces blood flow and causes capillary damage. The anthocyanins can scavenge these free radicals and prevents capillary damage (Bell and Gochenaur, 2006; Toufektsian et al., 2008).

4.6. Anthocyanins in vision improvement

Anthocyanin pigments maintain good eyes' health and are often associated with improving night vision (Kramer and Canter, 2004). Anthocyanin containing berries are beneficial for vision improvement in several ways, which includes (i) circulation increase within the retinal capillaries; (ii) improving night vision by the generation of retinal pigment; (iii) decreasing diabetic retinopathy and molecular degeneration; and (iv) preventing glaucoma, cataracts, and retinitis pigmentosa (Camire, 2000). Animal model studies show that anthocyanin-rich black currant and maqui berry have been used to improve eyesight (Matsumoto et al., 2006). Anthocyanins can be distributed through the blood-retinal and blood-aqueous barriers in ocular tissues. In 19 patients with openangle glaucoma, anthocyanin-rich black current (50 mg/day) supplementation improved ocular blood flow for 24 months (Ohguro et al., 2012). Some literature studies indicate that anthocyanin intake may positively impact macular degeneration associated with age and diabetic retinopathy (Nabavi et al., 2015; Yang et al., 2022). These results suggest that berries rich in anthocyanins could be used as a conventional treatment for patients suffering from open-angle glaucoma and other ophthalmic disorders.

4.7. Antimicrobial activity

Polyphenol compounds like anthocyanins show antimicrobial activity against several microorganisms, especially in inhibiting foodborne pathogens. Via different mechanisms, they exhibit antimicrobial activity, such as morphological damage to the bacterial cell or degradation of the structural integrity of the cell membrane, cell wall, and intracellular matrix (Burdulis et al., 2009). Anthocyanins, such as cyanidin, pelargonidin, and delphinidin, have been effective gram-negative inhibitors of Escherichia coli strain CM 871. However, these results were not reported in the case of wild variety gram-positive bacteria and E. coli (Nohynek et al., 2006). In addition to this, anthocyanins are responsible for plasma membrane permeabilization, extracellular microbial enzyme inhibition, and cytoplasmic membrane destabilization. Cranberries may inhibit various bacteria, including both gram-positive and gram-negative bacteria (Wu et al., 2008). Similarly, extract of blackcurrant inhibits the growth of Enterococcus faecium and Staphylococcus aureus strains, whereas it shows favorable results in the case of Saccharomyces cerevisiae and E. coli (Werlein et al., 2005). Likewise, carboxypyranocyanidin-3-O-glucoside from the anthocyanin family inhibits the biofilm production by S. aureus and Pseudomonas aeruginosa strains in chronic wounds (Correia et al., 2021). Despite all the evidence of anthocyanins showing antimicrobial activity, it has been shown that anthocyanins positively modulate the intestinal bacterial population by enhancing the growth of Lactobacillus enterococcus and Bifidobacterium spp., so the findings seem questionable in attributing the antimicrobial effects of the anthocyanin compounds observed (Hidalgo et al., 2012).

4.8. Antiviral activity

Anthocyanins have the potential to treat viral diseases. The structure of anthocyanin plays a vital role in restricting various viral infections. Blackberry extracts play a virucidal role against Herpes simplex due to a large number of anthocyanins. Due to the presence of anthocyanins in the red grape pomace extract, it reduces plaque infectivity and shows antiviral activity against H5N1 (Santhi et al., 2021). Similarly, Kannan and Kolandaivel (2018) concluded that the nature-derived cyanidin-3-sabubiocide could potentially treat the H1N1 subtype influenza virus. Likewise, red-fleshed potato contains Pg-type anthocyanins, which inactivates influenza viruses A and B (Hayashi et al., 2003). Study shows that anthocyanin-enriched elderberry fruit (Sambucus nigra L.) extract can demonstrate activity against modified vaccinia virus Ankara. It acts by declining the secretion of cytokinin IFN- γ and TNF- α , which promotes the Th2-Helper cell response (Schön et al., 2021). Anthocyanins from small red beans (Viqna angularis) can be used as a novel agent in the early stage of rabies virus infection (Kawai and Fujita, 2007). Anthocyanins have been recently explored as an alternative therapeutic agent targeting COVID-19. The ongoing COVID-19 pandemic, caused by severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2), has turned out a global health concern. It has affected millions of lives to date, and researchers are still hunting for its effective treatment. compounds cyanidin-3-O-β-glucoside Anthocvanin like (chrysanthemin) showed antiviral effects by binding to the naphthalene inhibitor binding site and inhibiting PLpro, though only to a limited extent. Similarly, pelargonidin, an anthocyanin, interact at a fatty acid-binding pocket on the spike RBD and inhibits spike-ACE2 interaction, lowering SARS-CoV-2 spike-ACE2 interaction and viral multiplication in Vero cells (Kaul et al., 2021). Furthermore, elderberry anthocyanins have the antiviral potential for SARS CoV-2 by limiting virus multiplication via budding from the virus's host cell (Salamon, 2020). A molecular docking simulation also suggests that berry anthocyanin could be employed as a potential SARS-CoV-2 therapy. The lowest binding energy, which was shown by cyanidin-3arabinoside, was found in the pocket through a sufficient number of hydrogen bonds with the major protease virus. Pelargonidin-3glucoside and pelargonidin 3-rhamnoside, on the other hand, have greater binding energy with SARS-spike CoV-2's glycoprotein (Messaoudi et al., 2021).

4.9. Other uses of anthocyanins

In food industries, color plays a major role, and the acceptability of food mostly depends upon its appearance. So many industries use synthetic color for improving elegance, neglecting its side effect. But the natural colorants derived from anthocyanins shows many beneficial effects without any adverse effect. Cyanidin, delphinidin, and pelargonidin are the anthocyanins responsible for various pigmentations (Mazza and Miniati, 2018). Anthocyanin extracts from bilberry and elderberry are accountable for producing mucus in the stomach and protecting it from ulcers (Veberic et al., 2009). A recent clinical trial demonstrated that the mixture of blue butterfly pea petals, extracts of purple waxy corn, and anthocyanin complex niosomes has a beneficial effect on oral wound healing (Damrongrungruang et al., 2021). Another study shows that cyanidin-3-glucoside from purple maize, fed to an obese mouse for 12 weeks, reduced body weight and reduced brown and white adipose tissue (Tsuda et al., 2003). The ability of anthocyanins to inhibit metalloproteases like MMP-1 and MMP-9 makes it suitable for topical application as they can protect the animal skin from ultraviolet radiation (Veberic et al., 2009). Additionally, anthocyanins from red cabbage possess hepatoprotective,

nephroprotective, hypocholesterolemic, and anti-obesity activity. These red cabbage anthocyanins also contain medicinal benefits against headaches, peptic ulcers, and gout (Ghareaghajlou et al., 2021). Apart from the above, much literature shows the beneficial effect of anthocyanins against diarrhea, hypertension, dysentery as analgesic and antifungal agents.

5. Conclusions

This review explains the current literature about anthocyanins' chemistry, absorption, and health benefits to give the reader a broad and updated scheme. It is believed that due to various pharmacological and beneficial activities, anthocyanins can be used as natural dietary sources in multiple diseases. Furthermore, the medical advantages of dietary anthocyanins have been shown in numerous *in vitro* and *in vivo* studies. Notably, it is evident from the above literature review that anthocyanins have been mainly explored in their extract form even with such medicinal properties. Therefore, it is time now that future studies should extensively focus on isolation and synthesis with the accurate and careful characterization of diverse anthocyanins as a sole chemical entity. This approach will allow the researchers to explore different mechanistic pathways about their biological effects and further intervene in their target-specific aspects

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Conflict of interest

The authors confirm that there are no known conflicts of interest.

CRediT authorship contribution statement

Sandip Patra: Resources, Data curation, Writing - original draft Priyanka N. Makhal: Investigation, Writing - Editing, Supervision Shubham Jaryal: Data curation, Writing - original draft Nilesh More: Data curation, Writing - original draft Venkata Rao Kaki: Conceptualization, Investigation, Supervision

ORCID Numbers of the Authors

S. Patra: 0000-0002-9679-3580 P.N. Makhal: 0000-0003-0833-8738 S. Jaryal: 0000-0001-5179-9519 N. More: 0000-0002-2189-2859 V.R. Kaki: 0000-0002-2840-861X

Supplementary File

None.

References

- Airoldi, C., La Ferla, B., D'Orazio, G., Ciaramelli, C., Palmioli, A., 2018. Flavonoids in the treatment of Alzheimer's and other neurodegenerative diseases. *Current Medicinal Chemistry*, 25(27), 3228-3246.
- Al-Awwadi, N.A., Araiz, C., Bornet, A., Delbosc, S., Cristol, J.P., Linck, N., Cros, G., 2005. Extracts enriched in different polyphenolic families normalize increased cardiac NADPH oxidase expression while having differential effects on insulin resistance, hypertension, and cardiac hypertrophy in high-fructose-fed rats. *Journal of Agricultural and Food Chemistry*, 53(1), 151-157.
- Ali, T., Kim, T., Rehman, S.U., Khan, M.S., Amin, F.U., Khan, M., Kim, M.O., 2018. Natural dietary supplementation of anthocyanins via PI3K/Akt/Nrf2/HO-1 pathways mitigate

oxidative stress, neurodegeneration, and memory impairment in a mouse model of Alzheimer's disease. *Molecular Neurobiology*, 55(7), 6076-6093.

- Alvarez-Suarez, J.M., Giampieri, F., Tulipani, S., Casoli, T., Di Stefano, G., González-Paramás, A.M., Battino, M., 2014. One-month strawberry-rich anthocyanin supplementation ameliorates cardiovascular risk, oxidative stress markers and platelet activation in humans. *The Journal of Nutritional Biochemistry*, 25(3), 289-294.
- Andriambeloson, E., Magnier, C., Haan-Archipoff, G., Lobstein, A., Anton, R., Beretz, A., Andriantsitohaina, R., 1998. Natural dietary polyphenolic compounds cause endothelium-dependent vasorelaxation in rat thoracic aorta. *The Journal of Nutrition*, 128(12), 2324-2333.
- Aqil, F., Gupta, A., Munagala, R., Jeyabalan, J., Kausar, H., Sharma, R.J., Gupta, R.C., 2012. Antioxidant and antiproliferative activities of anthocyanin/ellagitannin-enriched extracts from *Syzygium cumini* L.(Jamun, the Indian Blackberry). *Nutrition and Cancer*, 64(3), 428-438.
- Aqil, F., Vadhanam, M.V., Jeyabalan, J., Cai, J., Singh, I.P., upta, R.C., 2014. Detection of anthocyanins/anthocyanidins in animal tissues. *Journal of Agricultural and Food Chemistry*, 62(18), 3912-3918.
- Bell, D.R., Gochenaur, K., 2006. Direct vasoactive and vasoprotective properties of anthocyanin-rich extracts. *Journal of Applied Physiology*, 100(4), 1164-1170.
- Bensalem, J., Dal-Pan, A., Gillard, E., Calon, F., Pallet, V., 2015. Protective effects of berry polyphenols against age-related cognitive impairment. *Nutrition and Aging*, 3(2-4), 89-106.
- Bertuglia, S., Malandrino, S., Colantuoni, A., 1995. Effect of *Vaccinium myrtillus* anthocyanosides on ischaemia reperfusion injury in hamster cheek pouch microcirculation. *Pharmacological Research*, 31(3-4), 183-187.
- Blaut, M., Clavel, T., 2007. Metabolic diversity of the intestinal microbiota: implications for health and disease. *The Journal of Nutrition*, 137(3), 751S-755S.
- Borkowski, T., Szymusiak, H., Gliszczyńska-Świgło, A., Rietjens, I.M., Tyrakowska, B., 2005. Radical scavenging capacity of wine anthocyanins is strongly pH-dependent. *Journal of Agricultural and Food Chemistry*, 53(14), 5526-5534.
- Burdulis, D., Sarkinas, A., Jasutiene, I., Stackevicené, E., Nikolajevas, L., Janulis, V., 2009. Comparative study of anthocyanin composition, antimicrobial and antioxidant activity in bilberry (*Vaccinium myrtillus L.*) and blueberry (*Vaccinium corymbosum L.*) fruits. *Acta Poloniae Pharmaceutica*, 66(4), 399-408.
- Calderaro, A., Barreca, D., Bellocco, E., Smeriglio, A., Trombetta, D., Laganà, G., 2020. Colored phytonutrients: role and applications in the functional foods of anthocyanins. In Phytonutrients in Food (pp. 177-195). Woodhead Publishing.
- Camire, M.E., 2000. Bilberries and blueberries as functional foods and nutraceuticals. Functional Foods: Herbs, Botanicals and Teas. Lancaster: Technomic Publishing, pp. 289-319.
- Cao, H., Ou, J., Chen, L., Zhang, Y., Szkudelski, T., Delmas, D., Xiao, J., 2019. Dietary polyphenols and type 2 diabetes: Human Study and Clinical Trial. *Critical Reviews in Food Science and Nutrition*, 59(20), 3371-3379.
- Cao, W., Konsolaki, M., 2011. FKBP immunophilins and Alzheimer's disease: a chaperoned affair. *Journal of Biosciences*, 36(3), 493-498.
- Castañeda-Ovando, A., de Lourdes Pacheco-Hernández, M., Páez-Hernández, M.E., Rodríguez, J.A., Galán-Vidal, C.A., 2009. Chemical studies of anthocyanins: A review. *Food Chemistry*, 113(4), 859-871.
- Chen, Z., Wang, C., Pan, Y., Gao, X., Chen, H., 2018. Hypoglycemic and hypolipidemic effects of anthocyanins extract from black soybean seed coat in high fat diet and streptozotocin-induced diabetic mice. *Food & Function*, 9(1), 426-439.
- Clifford, M.N., 2000. Anthocyanins–nature, occurrence and dietary burden. Journal of the Science of Food and Agriculture, 80(7), 1063-1072.
- Correia, P., Araújo, P., Ribeiro, C., Oliveira, H., Pereira, A.R., Mateus, N., Fernandes, I., 2021. Anthocyanin-related pigments: Natural allies for skin health maintenance and protection. *Antioxidants*, 10(7), 1038.
- da Costa, C.T., Horton, D., Margolis, S.A., 2000. Analysis of anthocyanins in foods by liquid chromatography, liquid chromatography–mass spectrometry and capillary electrophoresis. *Journal of Chromatography A*, 881(1-2), 403-410.
- Damrongrungrung, T., Paphangkorakit, J., Limsitthichaikoon, S., Khampaenjiraroch, B., Davies, M.J., Sungthong, B., Priprem, A., 2021. Anthocyanin complex niosome gel accelerates oral wound healing: *In vitro* and clinical studies. *Nanomedicine: Nanotechnology, Biology and Medicine*, 37, 102423.
- De Ferrars, R.M., Czank, C., Zhang, Q., Botting, N.P., Kroon, P.A., Cassidy, A., Kay, C.D., 2014. The pharmacokinetics of anthocyanins and their metabolites in humans. *British Journal of Pharmacology*, 171(13), 3268-3282.
- Eder, R., 2000. Pigments. In Food analysis by HPLC (Second, pp. 825–880). Marcel Dekker Inc.
- El-Ella, D.M.A., Bishayee, A., 2019. The epigenetic targets of berry anthocyanins in cancer prevention. In Epigenetics of Cancer Prevention (pp. 129-148). Academic Press.
- Erlund, I., Koli, R., Alfthan, G., Marniemi, J., Puukka, P., Mustonen, P., Jula, A., 2008. Favorable effects of berry consumption on platelet function, blood pressure, and HDL cholesterol. *The American Journal of Clinical Nutrition*, 87(2), 323-331.
- Felgines, C., Texier, O., Besson, C., Lyan, B., Lamaison, J.L., Scalbert, A., 2007. Strawberry pelargonidin glycosides are excreted in urine as intact glycosides and glucuronidated pelargonidin derivatives in rats. *British Journal of Nutrition*, 98(6), 1126-1131.
- Fernandes, I., Faria, A., Calhau, C., de Freitas, V., Mateus, N., 2014. Bioavailability of anthocyanins and derivatives. *Journal of Functional Foods*, 7, 54-66.
- Forester, S.C., Choy, Y.Y., Waterhouse, A.L., Oteiza, P.I., 2014. The anthocyanin metabolites gallic acid, 3-O-methylgallic acid, and 2,4,6-trihydroxybenzaldehyde decrease human colon cancer cell viability by regulating pro-oncogenic signals. *Molecular Carcinogenesis*, 53(6), 432-439.

International Journal of Plant Based Pharmaceuticals, 2(1), 118-127

- Forester, S.C., Waterhouse, A.L., 2008. Identification of Cabernet Sauvignon anthocyanin gut microflora metabolites. *Journal of Agricultural and Food Chemistry*, 56(19), 9299-9304.
- Fossen, T., Cabrita, L., Andersen, O.M., 1998. Colour and stability of pure anthocyanins influenced by pH including the alkaline region. *Food Chemistry*, 63(4), 435-440.
- Frank, J., Kamal-Eldin, A., Lundh, T., Määttä, K., Törrönen, R., Vessby, B., 2002. Effects of dietary anthocyanins on tocopherols and lipids in rats. *Journal of Agricultural and Food Chemistry*, 50(25), 7226-7230.
- Garcia-Alonso, M., Minihane, A.M., Rimbach, G., Rivas-Gonzalo, J.C., de Pascual-Teresa, S., 2009. Red wine anthocyanins are rapidly absorbed in humans and affect monocyte chemoattractant protein 1 levels and antioxidant capacity of plasma. *The Journal of Nutritional Biochemistry*, 20(7), 521-529.
- Ghareaghajlou, N., Hallaj-Nezhadi, S., Ghasempour, Z., 2021. Red cabbage anthocyanins: Stability, extraction, biological activities and applications in food systems. *Food Chemistry*, 365, 130482.
- Giampieri, F., Gasparrini, M., Forbes-Hernandez, T.Y., Mazzoni, L., Capocasa, F., Sabbadini, S., Battino, M., 2018. Overexpression of the anthocyanidin synthase gene in strawberry enhances antioxidant capacity and cytotoxic effects on human hepatic cancer cells. *Journal of Agricultural and Food Chemistry*, 66(3), 581-592.
- Grimes, K.L., Stuart, C.M., McCarthy, J.J., Kaur, B., Cantu, E.J., Forester, S.C., 2018. Enhancing the cancer cell growth inhibitory effects of table grape anthocyanins. *Journal of Food Science*, 83(9), 2369-2374.
- Han, F., Yang, P., Wang, H., Fernandes, I., Mateus, N., Liu, Y., 2019. Digestion and absorption of red grape and wine anthocyanins through the gastrointestinal tract. *Trends in Food Science & Technology*, 83, 211-224.
- Hanske, L., Engst, W., Loh, G., Sczesny, S., Blaut, M., Braune, A., 2013. Contribution of gut bacteria to the metabolism of cyanidin 3-glucoside in human microbiotaassociated rats. *British Journal of Nutrition*, 109(8), 1433-1441.
- Hayashi, K., Mori, M., Knox, Y. M., Suzutan, T., Ogasawara, M., Yoshida, I., Azuma, M., 2003. Anti influenza virus activity of a red-fleshed potato anthocyanin. *Food Science* and Technology Research, 9(3), 242-244.
- He, J., Giusti, M.M., 2010. Anthocyanins: natural colorants with health-promoting properties. Annual Review of Food Science and Technology, 1, 163-187.
- He, J., Wallace, T.C., Keatley, K.E., Failla, M.L., Giusti, M.M., 2009. Stability of black raspberry anthocyanins in the digestive tract lumen and transport efficiency into gastric and small intestinal tissues in the rat. *Journal of Agricultural and Food Chemistry*, 57(8), 3141-3148.
- Hidalgo, M., Oruna-Concha, M.J., Kolida, S., Walton, G.E., Kallithraka, S., Spencer, J.P., de Pascual-Teresa, S., 2012. Metabolism of anthocyanins by human gut microflora and their influence on gut bacterial growth. *Journal of Agricultural and Food Chemistry*, 60(15), 3882-3890.
- Hossain, M.K., Abdal Dayem, A., Han, J., Yin, Y., Kim, K., Kumar Saha, S., Cho, S.G., 2016. Molecular mechanisms of the anti-obesity and anti-diabetic properties of flavonoids. *International Journal of Molecular Sciences*, 17(4), 569.
- Hou, D.X., 2003. Potential mechanisms of cancer chemoprevention by anthocyanins. *Current Molecular Medicine*, 3(2), 149-159.
- Hribar, U., Poklar Ulrih, N., 2014. The metabolism of anthocyanins. Current Drug Metabolism, 15(1), 3-13.
- Igarashi, K., Kimura, Y., Takenaka, A., 2000. Preventive effects of dietary cabbage acylated anthocyanins on paraquat-induced oxidative stress in rats. *Bioscience*, *Biotechnology*, and *Biochemistry*, 64(8), 1600-1607.
- Iversen, C., 1996. Degradation of C-vitamin and anthocyanins in blackcurrant juice. In Proceedings of the Symposium on Polyphenols and Anthocyanins as Food Colourants and Antioxidants, Ed by Pfannhauser W and Strigl A. EU, Brussels, Belgium, pp 34.
- Jaganathan, S.K., Vellayappan, M.V., Narasimhan, G., Supriyanto, E., Dewi, D.E.O., Narayanan, A.L.T., Yusof, M., 2014. Chemopreventive effect of apple and berry fruits against colon cancer. *World Journal of Gastroenterology*, 20(45), 17029–17036.
- Janeiro, P., Brett, A.M.O., 2007. Redox behavior of anthocyanins present in Vitis vinifera L. Electroanalysis: An International Journal Devoted to Fundamental and Practical Aspects of Electroanalysis, 19(17), 1779-1786.
- Jang, I.S., Kim, D.H., 1996. Purification and characterization of α -L-rhamnosidase from Bacteroides JY-6, a human intestinal bacterium. *Biological and Pharmaceutical Bulletin*, 19(12), 1546-1549.
- Joshi, R., Rana, A., Kumar, V., Kumar, D., Padwad, Y.S., Yadav, S.K., Gulati, A., 2017. Anthocyanins enriched purple tea exhibits antioxidant, immunostimulatory and anticancer activities. *Journal of Food Science and Technology*, 54(7), 1953-1963.
- Jung, U.J., Kim, S.R., 2018. Beneficial effects of flavonoids against Parkinson's disease. Journal of Medicinal Food, 21(5), 421-432.
- Kannan, S., Kolandaivel, P., 2018. The inhibitory performance of flavonoid cyanidin-3sambubiocide against H274Y mutation in H1N1 influenza virus. *Journal of Biomolecular Structure and Dynamics*, 36(16), 4255-4269.
- Kaul, R., Paul, P., Kumar, S., Büsselberg, D., Dwivedi, V D., Chaari, A., 2021. Promising antiviral activities of natural flavonoids against SARS-CoV-2 targets: systematic review. *International Journal of Molecular Sciences*, 22(20), 11069.
- Kawai, A., Fujita, K., 2007. Small red bean (azuki) sheds biologically active substances as a prerequisite step for germination, one of which displays the antiviral activity against the rabies virus infectivity and infections in culture. *Microbiology and Immunology*, 51(11), 1071-1079.
- Kay, C.D., 2006. Aspects of anthocyanin absorption, metabolism and pharmacokinetics in humans. *Nutrition Research Reviews*, 19(1), 137-146.
- Keppler, K., Humpf, H.U., 2005. Metabolism of anthocyanins and their phenolic degradation products by the intestinal microflora. *Bioorganic & Medicinal Chemistry*, 13(17), 5195-5205.

International Journal of Plant Based Pharmaceuticals, 2(1), 118-127

- Khan, M.S., Ali, T., Kim, M.W., Jo, M.H., Chung, J.I., Kim, M.O., 2019. Anthocyanins improve hippocampus-dependent memory function and prevent neurodegeneration via JNK/Akt/GSK3β signaling in LPS-treated adult mice. *Molecular Neurobiology*, 56(1), 671-687.
- Khanra, R., Dewanjee, S., Dua, T.K., Sahu, R., Gangopadhyay, M., De Feo, V., Zia-Ul-Haq, M., 2015. Abroma augusta L. (Malvaceae) leaf extract attenuates diabetes induced nephropathy and cardiomyopathy via inhibition of oxidative stress and inflammatory response. Journal of Translational Medicine, 13(1), 1-14.
- Khoo, H.E., Azlan, A., Tang, S.T., Lim, S.M., 2017. Anthocyanidins and anthocyanins: colored pigments as food, pharmaceutical ingredients, and the potential health benefits. *Food & Nutrition Research*, 61(1), 1361779.
- Kong, J.M., Chia, L.S., Goh, N.K., Chia, T.F., Brouillard, R., 2003. Analysis and biological activities of anthocyanins. *Phytochemistry*, 64(5), 923-933.
- Kramer, J.H., Canter, P.H., 2004. Anthocyanosides of Vaccinium myrtillus (bilberry) for night vision—a systematic review of placebo-controlled trials. Survey of Ophthalmology, 49(6), 618.
- Krga, I., Milenkovic, D., 2019. Anthocyanins: From sources and bioavailability to cardiovascular-health benefits and molecular mechanisms of action. *Journal of Agricultural and Food Chemistry*, 67(7), 1771-1783.
- Leong, H.Y., Show, P.L., Lim, M.H., Ooi, C.W., Ling, T.C., 2018. Natural red pigments from plants and their health benefits: A review. *Food Reviews International*, 34(5), 463-482.
- Li, D., Wang, P., Luo, Y., Zhao, M., Chen, F., 2017. Health benefits of anthocyanins and molecular mechanisms: Update from recent decade. *Critical Reviews in Food Science* and Nutrition, 57(8), 1729-1741.
- Li, H., Deng, Z., Zhu, H., Hu, C., Liu, R., Young, J.C., Tsao, R., 2012. Highly pigmented vegetables: Anthocyanin compositions and their role in antioxidant activities. *Food Research International*, 46(1), 250-259.
- Lim, S., Xu, J., Kim, J., Chen, T.Y., Su, X., Standard, J., Wang, W., 2013. Role of anthocyanin-enriched purple-fleshed sweet potato p40 in colorectal cancer prevention. *Molecular Nutrition & Food Research*, 57(11), 1908-1917.
- Lin, B.W., Gong, C.C., Song, H.F., Cui, Y.Y., 2017. Effects of anthocyanins on the prevention and treatment of cancer. *British Journal of Pharmacology*, 174(11), 1226-1243.
- Lo Piero, A.R., 2015. The state of the art in biosynthesis of anthocyanins and its regulation in pigmented sweet oranges [(*Citrus sinensis*) L. Osbeck]. *Journal of Agricultural and Food Chemistry*, 63(16), 4031-4041.
- Lucioli, S., 2012. Anthocyanins: Mechanism of action and therapeutic efficacy. In Medicinal Plants as Antioxidant Agents: Understanding Their Mechanism of Action and Therapeutic Efficacy (Vol. 661, Issue 2, pp. 27–57). Research Signpost Kerala, India.
- Maestro, A., Terdoslavich, M., Vanzo, A., Kuku, A., Tramer, F., Nicolin, V., Passamonti, S., 2010. Expression of bilitranslocase in the vascular endothelium and its function as a flavonoid transporter. *Cardiovascular Research*, 85(1), 175-183.
- Marquart, L.C., 1835. Die Farben der Blüthen: Eine chemisch-physiolog. Abhandlung. Habicht.
- Matsumoto, H., Nakamura, Y., lida, H., Ito, K., Ohguro, H., 2006. Comparative assessment of distribution of blackcurrant anthocyanins in rabbit and rat ocular tissues. *Experimental Eye Research*, 83(2), 348-356.
- Mazewski, C., Liang, K., de Mejia, E.G., 2018. Comparison of the effect of chemical composition of anthocyanin-rich plant extracts on colon cancer cell proliferation and their potential mechanism of action using *in vitro*, *in silico*, and biochemical assays. *Food Chemistry*, 242, 378-388.
- Mazza, G., Miniati, E., 2018. Anthocyanins in fruits, vegetables, and grains. CRC press, (vol. 362).
- McGhie, T.K., Walton, M.C., 2007. The bioavailability and absorption of anthocyanins: towards a better understanding. Molecular Nutrition & Food Research, 51(6), 702-713.
- Messaoudi, O., Gouzi, H., El-Hoshoudy, A.N., Benaceur, F., Patel, C., Goswami, D., Bendahou, M., 2021. Berries anthocyanins as potential SARS-CoV–2 inhibitors targeting the viral attachment and replication; molecular docking simulation. *Egyptian Journal of Petroleum*, 30(1), 33-43.
- Moshfegh, F., Balanejad, S.Z., Shahrokhabady, K., Attaranzadeh, A., 2022. *Crocus sativus* (saffron) petals extract and its active ingredient, anthocyanin improves ovarian dysfunction, regulation of inflammatory genes and antioxidant factors in testosterone-induced PCOS mice. *Journal of Ethnopharmacology*, 282, 114594.
- Muñoz-González, I., Jiménez-Girón, A., Martín-Álvarez, P.J., Bartolomé, B., Moreno-Arribas, M.V., 2013. Profiling of microbial-derived phenolic metabolites in human feces after moderate red wine intake. *Journal of Agricultural and Food Chemistry*, 61(39), 9470-9479.
- Murapa, P., Dai, J., Chung, M., Mumper, R.J., D'Orazio, J., 2012. Anthocyanin-rich fractions of blackberry extracts reduce UV-induced free radicals and oxidative damage in keratinocytes. *Phytotherapy Research*, 26(1), 106-112.
- Nabavi, S.F., Habtemariam, S., Daglia, M., Shafighi, N., Barber, A.J., Nabavi, S.M., 2015. Anthocyanins as a potential therapy for diabetic retinopathy. *Current Medicinal Chemistry*, 22(1), 51-58.
- Nohynek, L.J., Alakomi, H.L., Kähkönen, M.P., Heinonen, M., Helander, I.M., Oksman-Caldentey, K.M., Puupponen-Pimiä, R.H., 2006. Berry phenolics: antimicrobial properties and mechanisms of action against severe human pathogens. *Nutrition and Cancer*, 54(1), 18-32.
- Nurmi, T., Mursu, J., Heinonen, M., Nurmi, A., Hiltunen, R., Voutilainen, S., 2009. Metabolism of berry anthocyanins to phenolic acids in humans. *Journal of Agricultural* and Food Chemistry, 57(6), 2274-2281.

- Ohguro, H., Ohguro, I., Katai, M., Tanaka, S., 2012. Two-year randomized, placebocontrolled study of black currant anthocyanins on visual field in glaucoma. *Ophthalmologica*, 228(1), 26-35.
- Pan, Z., Cui, M., Dai, G., Yuan, T., Li, Y., Ji, T., Pan, Y., 2018. Protective effect of anthocyanin on neurovascular unit in cerebral ischemia/reperfusion injury in rats. *Frontiers in Neuroscience*, 947.
- Passamonti, S., Vrhovsek, U., Mattivi, F., 2002. The interaction of anthocyanins with bilitranslocase. *Biochemical and Biophysical Research Communications*, 296(3), 631-636.
- Passamonti, S., Vrhovsek, U., Vanzo, A., Mattivi, F., 2003. The stomach as a site for anthocyanins absorption from food. *FEBS Letters*, 544(1-3), 210-213.
- Pesce, C., Menini, S., 2019. A Protocol Outline of Dietary Intervention to Contrast Diabetic Nephropathy. In Bioactive Food as Dietary Interventions for Diabetes (pp. 33-48). Academic Press.
- Pina, F., Melo, M. J., Laia, C.A., Parola, A.J., Lima, J.C., 2012. Chemistry and applications of flavylium compounds: a handful of colours. *Chemical Society Reviews*, 41(2), 869-908.
- Pojer, E., Mattivi, F., Johnson, D., Stockley, C.S., 2013. The case for anthocyanin consumption to promote human health: a review. *Comprehensive Reviews in Food Science and Food Safety*, 12(5), 483-508.
- Prior, R.L., 2004. Absorption and metabolism of anthocyanins: potential health effects. In A. J. Meskin, M.S., Bidlack, W.R., Davies (Ed.), Phytochemicals: Mechanisms of Action, (1st Edition, pp. 1–19). CRC Press.
- Prior, R.L., Wu, X., 2006. Anthocyanins: structural characteristics that result in unique metabolic patterns and biological activities. *Free Radical Research*, 40(10), 1014-1028.
- Rasheed, Z., Akhtar, N., Anbazhagan, A.N., Ramamurthy, S., Shukla, M., Haqqi, T.M., 2009. Polyphenol-rich pomegranate fruit extract (POMx) suppresses PMACI-induced expression of pro-inflammatory cytokines by inhibiting the activation of MAP Kinases and NF-κB in human KU812 cells. *Journal of Inflammation*, 6(1), 1-12.
- Richardson, P.M., Harborne, J.B., 1990. Phytochemical Methods: A Guide to Modern Techniques of Plant Analysis. Second Edition. In Brittonia (Vol. 42, Issue 2). Springer Science & Business Media.
- Roy, M., Sen, S., Chakraborti, A.S., 2008. Action of pelargonidin on hyperglycemia and oxidative damage in diabetic rats: implication for glycation-induced hemoglobin modification. *Life Sciences*, 82(21-22), 1102-1110.
- Salamon, I, 2020. Can Medicinal Plants Help in the Treatment of the New Coronavirus? Some R & D Aspects in Slovak Republic. *Current Perspectives on Medicinal and Aromatic Plants (CUPMAP)*, 4(1), 58-65.
- Santhi, V.P., Sriramavaratharajan, V., Murugan, R., Masilamani, P., Gurav, S.S., Sarasu, V.P., Ayyanar, M., 2021. Edible fruit extracts and fruit juices as potential source of antiviral agents: a review. *Journal of Food Measurement and Characterization*, 15(6), 5181-5190.
- Schön, C., Mödinger, Y., Krüger, F., Doebis, C., Pischel, I., Bonnländer, B., 2021. A new high-quality elderberry plant extract exerts antiviral and immunomodulatory effects in vitro and ex vivo. Food and Agricultural Immunology, 32(1), 650-662.
- Shah, S.A., Amin, F.U., Khan, M., Abid, M.N., Rehman, S.U., Kim, T.H., Kim, M.O., 2016. Anthocyanins abrogate glutamate-induced AMPK activation, oxidative stress, neuroinflammation, and neurodegeneration in postnatal rat brain. *Journal of Neuroinflammation*, 13(1), 1-16.
- Shaik, A., Naidu, K.K., Panda, J., 2018. A review on anthocyanins: a promising role on phytochemistry and pharmacology. *International Research Journal of Pharmacy*, 9(1), 1-9.
- Shih, P.H., Wu, C.H., Yeh, C.T., Yen, G.C., 2011. Protective effects of anthocyanins against amyloid β-peptide-induced damage in neuro-2A cells. *Journal of Agricultural* and Food Chemistry, 59(5), 1683-1689.
- Singletary, K.W., Jung, K.J., Giusti, M., 2007. Anthocyanin-rich grape extract blocks breast cell DNA damage. *Journal of Medicinal Food*, 10(2), 244-251.
- Smeriglio, A., Barreca, D., Bellocco, E., Trombetta, D., 2016. Chemistry, pharmacology and health benefits of anthocyanins. *Phytotherapy Research*, 30(8), 1265-1286.
- Sonmez, F., Gunesli, Z., Demir, T., Cıkrıkcı, K., Ergun, A., Gencer, N., Arslan, O., 2022. The Effect of Total Anthocyanins Extracted From Sweet Cherry Cultivars on Carbonic Anhydrases and Antioxidant Activity. Erwerbs-Obstbau, 1-9.
- Stalmach, A., Edwards, C.A., Wightman, J.D., Crozier, A., 2013. Colonic catabolism of dietary phenolic and polyphenolic compounds from Concord grape juice. *Food & Function*, 4(1), 52-62.
- Talavéra, S., Felgines, C., Texier, O., Besson, C., Manach, C., Lamaison, J.L., Rémésy, C., 2004. Anthocyanins are efficiently absorbed from the small intestine in rats. *The Journal of Nutrition*, 134(9), 2275-2279.
- Tarozzi, A., Morroni, F., Merlicco, A., Bolondi, C., Teti, G., Falconi, M., Hrelia, P., 2010. Neuroprotective effects of cyanidin 3-O-glucopyranoside on amyloid beta (25-35) oligomer-induced toxicity. *Neuroscience Letters*, 473(2), 72-76.
- Timberlake, C.F., Henry, B.S., 1988. Anthocyanins as natural food colorants. *Progress in Clinical and Biological Research*, 280, 107-121.
- Toufektsian, M.C., De Lorgeril, M., Nagy, N., Salen, P., Donati, M.B., Giordano, L., Martin, C., 2008. Chronic dietary intake of plant-derived anthocyanins protects the rat heart against ischemia-reperfusion injury. *The Journal of Nutrition*, 138(4), 747-752.
- Tsuda, T., Horio, F., Uchida, K., Aoki, H., Osawa, T., 2003. Dietary cyanidin 3-*O*-β-Dglucoside-rich purple corn color prevents obesity and ameliorates hyperglycemia in mice. *The Journal of Nutrition*, 133(7), 2125-2130.
- Tsuda, T., Shiga, K., Ohshima, K., Kawakishi, S., Osawa, T., 1996. Inhibition of lipid peroxidation and the active oxygen radical scavenging effect of anthocyanin pigments isolated from *Phaseolus vulgaris* L. *Biochemical Pharmacology*, 52(7), 1033-1039.

- Turturică, M., Oancea, A.M., Râpeanu, G., Bahrim, G., 2015. Anthocyanins: naturally occuring fruit pigments with functional properties. *Annals of the University Dunarea de Jos of Galati Fascicle VI--Food Technology*, 39(1), 9–24.
- Vanamala, J.K.P., 2019. Potatoes for targeting colon cancer stem cells. *American Journal of Potato Research*, 96(2), 177-182.
- Veberic, R., Jakopic, J., Stampar, F., Schmitzer, V., 2009. European elderberry (Sambucus nigra L.) rich in sugars, organic acids, anthocyanins and selected polyphenols. Food Chemistry, 114(2), 511-515.
- Wang, L.S., Stoner, G.D., 2008. Anthocyanins and their role in cancer prevention. *Cancer Letters*, 269(2), 281-290.
- Wang, S.Y., Jiao, H., 2000. Scavenging capacity of berry crops on superoxide radicals, hydrogen peroxide, hydroxyl radicals, and singlet oxygen. *Journal of Agricultural and Food Chemistry*, 48(11), 5677-5684.
- Werlein, H.D., Kütemeyer, C., Schatton, G., Hubbermann, E.M., Schwarz, K., 2005. Influence of elderberry and blackcurrant concentrates on the growth of microorganisms. *Food Control*, 16(8), 729-733.
- Wu, V.C.H., Qiu, X., Bushway, A., Harper, L., 2008. Antibacterial effects of American cranberry (Vaccinium macrocarpon) concentrate on foodborne pathogens. LWT-Food Science and Technology, 41(10), 1834-1841.
- Yang, M., Koo, S.I., Song, W.O., Chun, O.K., 2011. Food matrix affecting anthocyanin bioavailability. *Current Medicinal Chemistry*, 18(2), 291-300.
- Yang, W., Guo, Y., Liu, M., Chen, X., Xiao, X., Wang, S., Chen, F., 2022. Structure and function of blueberry anthocyanins: A review of recent advances. *Journal of Functional Foods*, 88, 104864.
- Zafra-Stone, S., Yasmin, T., Bagchi, M., Chatterjee, A., Vinson, J.A., Bagchi, D., 2007. Berry anthocyanins as novel antioxidants in human health and disease prevention. *Molecular Nutrition & Food Research*, 51(6), 675-683.
- Zhang, H., Wang, L., Deroles, S., Bennett, R., Davies, K., 2006. New insight into the structures and formation of anthocyanic vacuolar inclusions in flower petals. *BMC Plant Biology*, 6(1), 1-14.
- Zhang, J., Xiao, J., Giampieri, F., Forbes-Hernandez, T.Y., Gasparrini, M., Afrin, S., Cianciosi, D., Reboredo-Rodriguez, P., Battino, M., Zheng, X., 2019. Inhibitory effects of anthocyanins on α-glucosidase activity. *Journal of Berry Research*, 9(1), 109–123.
- Zhang, Y., Seeram, N.P., Lee, R., Feng, L., Heber, D., 2008. Isolation and identification of strawberry phenolics with antioxidant and human cancer cell antiproliferative properties. *Journal of Agricultural and Food Chemistry*, 56(3), 670-675.
- Zhang, Y., Vareed, S.K., Nair, M.G., 2005. Human tumor cell growth inhibition by nontoxic anthocyanidins, the pigments in fruits and vegetables. *Life Sciences*, 76(13), 1465-1472.

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Zhu, W., Jia, Q., Wang, Y., Zhang, Y., Xia, M., 2012. The anthocyanin cyanidin-3-*O*-βglucoside, a flavonoid, increases hepatic glutathione synthesis and protects hepatocytes against reactive oxygen species during hyperglycemia: Involvement of a cAMP–PKA-dependent signaling pathway. *Free Radical Biology and Medicine*, 52(2), 314-327.

Reviewed by:

Kagolla PRISCILLA: Central University of Karnataka, Kadaganchi, INDIA

Mostapha BACHIR-BEY: University of Bejaia, Bejaia, ALGERIA

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